1 Contextualising samples: Supporting reference genomes

² for <u>of</u> European biodiversity through sample and

³ associated metadata collection

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90 Abstract

91 The European Reference Genome Atlas (ERGA) consortium aims to generate a reference 92 genome catalogue for all of Europe's eukaryotic biodiversity. The biological material underlying this mission, the specimens and their derived samples, are provided through ERGA's pan-93 94 European network. To demonstrate the community's capability and capacity to realise ERGA's 95 ambitious mission, the ERGA Pilot project was initiated. In support of the ERGA Pilot effort to 96 generate reference genomes for European biodiversity, the ERGA Sampling and Sample 97 Processing committee (SSP) was formed by volunteer experts from ERGA's member base. SSP 98 aims to aid participating researchers through i) establishing standards for and collecting of 99 sample/ specimen metadata; ii) prioritisation of species for genome sequencing; and iii) 100 development of taxon-specific collection guidelines including logistics support. SSP serves as 101 the sample provider's entry point for samplesamples providers to the ERGA genomic resource 102 production infrastructure and guarantees that ERGA's high-guality standards are upheld 103 throughout sample collection and processing. With the volume of researchers, projects, 104 consortia, and organisations with interests in genomics resources expanding, this manuscript 105 shares important experiences and lessons learned during the development of standardised 106 operational procedures and sample provider support. The manuscript details our experiences in 107 incorporating the FAIR and CARE principles, species prioritisation, and workflow development, 108 which could be useful to individuals as well as other initiatives.

I. The Sampling and Sample Processing committee ofERGA

111 The European Reference Genome Atlas (ERGA, Mazzoni et al. 2023) consortium, the European 112 node of the Earth BioGenome Project (EBP; Lewin et al. 2022), aims to generate a publicly 113 available reference genome catalogue for all European eukarvotic biodiversity (Formenti et al. 114 2022; Theissinger et al. 2023). ERGA has the potential to catapult the fields of biodiversity 115 conservation, evolution, ecology, and others to a new sphere analogous to how the first complete 116 sequence of the human genome surged the fields of medical genetics, genomics, anthropology, 117 and others (Formenti et al. 2022; Theissinger et al. 2023). It is akin to the appearance of the first 118 natural history collections dating back as far as the 1800s that still lay the foundations for many 119 new and important insights today.

120 ERGA is led by its chair and two co-chairs in cooperation with the ERGA council (a team 121 consisting of two elected representatives of each member country). To support the multitude of 122 ERGA tasks, several scientific and Science+ committees have been established, one of which 123 is the Sampling and Sample Processing committee (SSP). ERGA's first project - the ERGA Pilot 124 (McCartney et al. 2023), tested a distributed genomics infrastructure while fuelling the ERGA 125 committees. The Pilot Project is a community effort without a dedicated funding source, which 126 will result in the production of over 98 genomes from 34 provider countries, connecting close to 127 400 involved ERGA members.

The Sampling and Sample Processing committee (SSP)SSP is a working-committeegroup of volunteer expert ERGA members tasked with developing guidelines to support sampling and sample processing. Specifically, the SSP's initial responsibilities included i) establishing standards and mechanisms to collect sample/specimen metadata; ii) prioritising species collection; and iii) developing taxon-specific collection guidelines for the biological material underlying ERGA's mission. The specimens and their derived samples are provided through ERGA's large network of biodiversity partners spread across Europe (Box 1).

- The SSP serves as the sample provider's entry point into ERGA's distributed genomic infrastructure and helps ensure standardised sample processing. As ERGA was maturing, additional SSP tasks emerged: iv) providing guidance to sample providers for the compliance with legal obligations in collaboration with ERGA's <u>ELSI committee</u> (<u>Ethical, Legal, and Social</u> <u>Issues</u>) and v) sample provision - facilitating sample shipping between sample providers and sequencing centres.
- 141 As the number of EBP-associated projects across the globe gradually increases, we share here 142 the experiences we gained whilst developing the operational procedures and sample provider 143 support systems for the first continent-wide, distributed, genomics infrastructure. We hope our 144 lessons can be useful to other large consortia who are pursuing the shared mission of 145 sequencing all of life. Our experience in tackling FAIR (Findable, Accessible, Interoperable, 146 Reusable) and CARE (Collective benefit, Authority to control, Responsibility, Ethics) data 147 principles, species prioritisation, and workflow development may also be of use to smaller 148 initiatives.

Box1. The scheme shows the ERGA workflow in the Pilot project. Species were initially nominated by the ERGA community (1), accompanied by a comprehensive form containing questions used for Species Selection (2), based on several exclusion, prioritisation and feasibility criteria. Species were distributed to the participating Sequencing Partners (3), which were responsible to contact the Genome Team lead (often the sample provider) to organise all necessary onboarding and regulatory requirements and documentation and agreed to generate reference genomes that fulfil <u>EBP</u> <u>quality metrics</u> (4). Samples were collected, vouchered, and several tubes of subsamples were prepared for sequencing as arranged with the sequencing partner and collaborating research groups (5). Sample providers were also encouraged to barcode the samples prior to sequencing and to store corresponding material in local biobanking facilities. Metadata was recorded using the ERGA sample manifest following established guidelines (6), uploaded to the metadata brokering platform COPO and validated by the Pilot sample management team (7). After confirmation that all the required documentation and metadata was in place, samples were shipped assuring a cold chain to the designated sequencing facility (8).

Species Nominations 2 Species Selection Prioritization Criteria Prioritization Criteria Nomination Scientific
Assignment to Sequencing Center According to: • Capacity to generate the data volumes to fulfil EBP standards for genome assembly • Sample requirements.
C. are in place. Those of a point of the following series and the
Collect the samples, following the requirements specified from the assigned Sequencing Facility. Collect the speciment following the taxon/sequencing facility specific standards) and take scaled pictures. Collect the speciments following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate a speciment following the taxon/sequencing facility specific guidelines, and separate speciment fol
Barcode the samples prior to sample Barcode the samples prior to samples prior to samples Barcode the samples pr
Cell culture (optional) : If planning to send samples for Cell culture, please follow these instructions, and get in touch with <u>ERGA-cells@upf_edu</u> for guidance.
Betadata recording and Sample Manifest A Sample Manifest Upload Set up an ORCID account and contact COPO team guidelines. Set up an ORCID account and contact COPO team Set up an ORCID account and contact COPO team
Note: Select REFERENCE_GENOME" from the drop- down menu on column C "PURPOSE_OF_SPECIMEN" of the Manifest Upload the ERGA Sample Manifest to <u>COPO</u> . following the provided guidelines. The Sample management team will validate the Sample Manifest and give the green light to start the shipping process in coordination with the Sequencing Centre.
8 Sample Shipping All samples must be shipped on dry ice or in a dry shipper .

150 Reference genome production within a multinational consortium like ERGA involves many 151 partners spanning dozens of countries. To manage diverse expectations, ensure efficient task 152 execution, streamline communication, and safeguard fair attribution, ERGA has implemented 153 the formation of multidisciplinary 'Genome Teams' (Supplementary File 1). These include all 154 contributors to the production of a reference genome (i.e., researchers, stakeholders, and rights 155 holders) from the field to the final data analysis. The Genome Team lead's (in the ERGA Pilot 156 known as the sample ambassador) initial responsibilities include providing all necessary 157 documentation, data, and metadata for a sample to enter the sequencing workflow (Box 1). Most 158 often, this function is filled by the sample provider. All members of the Genome Team agree to 159 adhere to ERGA's Sample Code of Practice as well as ERGA's Code of Conduct. The SSP 160 committee serves as an important touch point for the Genome Team lead, providing advice and 161 guidance on sampling requirements, metadata standards, legal compliance, and vouchering 162 strategies.

Selecting species for biodiversity genomics - species prioritisation 163 in ERGA's initial phase 164

165 Reference genome sequencing initiatives require implementing prioritisation criteria, given 166 resource and technical limitations that prevent sequencing all targeted species immediately. 167 Scientific, technical, and social criteria can govern such species prioritisation.

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Criteria	Scientific criteria	Technical criteria	Social criteria
Examples	taxonomic representation/targets	sample availability including voucher specimen	importance to local communities
	conservation status	specimen/sample size (amount of biological material and therefore DNA and/or RNA)	cultural significance
	value of genome for specific field of interest (e.g., biomedicine, biotechnology, agriculture)	sampling and handling logistics	inclusiveness targets concerning countries and individuals
	Taxonomic certainty	genome characteristics (estimated genome size and ploidy)	community engagement

169 Table 1 Non-exhaustive list of criteria for species prioritisation for genome sequencing projects

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172 For initiating ERGA as a continent-wide genomic infrastructure network, a pool of candidate 173 species for reference genome generation was solicited that were representative of the diversity

174 of species and scientists across the consortium. To this aim, the ERGA community was asked 175 to propose species through an initial simple ERGA species suggestion form resulting in 276

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nominations. Subsequently, nominating persons were contacted to complete a comprehensive

form (Supplementary File 2) containing 117 questions and commenting fields. The form included questions related to taxonomic identity, genome properties, voucher availability, habitat of species in question, sampling strategy, species conservation status, permits to obtain material for genome sequencing, sample properties (e.g., sex, amount, preservation quality, and tissue type), and species identification certainty. The refined species nomination form was open for 26 days and received 155 submissions.

183 After excluding species that already had available reference genomes, SSP implemented a 184 prioritisation process based on country of origin and a simple scoring system, attributing a score 185 of 1 to 3 in eight categories (Table 2). Higher priority was given to species that: i) had a genome 186 size smaller than 1Gb, ii) were readily available, iii) could be freshly collected and for which 187 biological material could be flash frozen, iv) could deliver >1g of tissue (if the organism permitted) 188 and had well-established extraction protocols that allowed isolating chemically pure HMW DNA. 189 v) could deposit a specimen voucher, vi) had no ambiguity risk in species identification, vii) had 190 all permits present or were not needed (a formal documentation for either of the solutions was

191 requested), and viii) had no export restrictions (if applicable).

192 After ranking the species according to this scoring system, each proposing country was given 193 the opportunity to refine their selection of species and to propose three final species considering 194 three predefined target categories (endangered/iconic, marine/freshwater and pollinator) to 195 match the available resources. At that point, ERGA had no centralised funding so feasibility was 196 strongly determined by the availability of sufficient funds to support genome sequencing for a 197 particular species. The project relied on resources contributed by participating ERGA members, 198 institutions, and sequencing centres, with some additional support from industrial sponsors, that 199 was used to supplement equity deserving genome teams in order to improve wide access to 200 participation. As an extension to the selected list, standalone species were also included under 201 the ERGA umbrella if they were completely funded by independent resources.

The circulation of the list of nominated species within ERGA resulted in cross-country collaborations especially for species proposed by more than one country, fostering exchange and reducing costs and redundancies.

The species selection and prioritisation process resulted in 98 selected species (<u>https://goat.genomehubs.org/projects/ERGA</u>), from 15 phyla (<u>Figure 1B</u>) and 34 countries or regions (Figure 1). With six of the seven selection scores relating to feasibility (including legal), this was the most prominent criterion, while the other criteria (i.e., conservation status, scientific relevance, socioeconomic relevance, taxonomic gaps, and community engagement) played only an indirect role via the subjective selection by the ERGA council members. <u>ERGA has planned</u> to implement unbiased species selection procedures in the future to alleviate the dominance of

212 <u>feasibility as selection criterion (see section V below).</u>

Both the initial and the final list of selected species showed a predominance of chordates, arthropods, and tracheophytes. Given that the initial pool of species was suggested by the ERGA community, this predominance may reflect the organism-bias of the biodiversity genomics community at large (see below). This taxon bias remained despite the dynamic nature of the taxonomic composition, as some species were removed due to sampling or sequencing 218 technical barriers whilst others were added to increase representation and participation across 219 ERGA's diverse members. A total of 37% of the species were considered for the category 220 endangered/iconic, and 12% were pollinators (as one example of scientific relevance and a 221 target group of the Biodiversity Strategy of the European Commission). Most of the reference 222 genomes were generated because the species are endemic (28%), endangered (26%) (and 223 therefore the genome could be leveraged to inform conservation plans in the future) or to be 224 used to answer specific scientific questionsfor research purposes (25%) (Figure 1C), and tThe 225 most popular planned downstream analyses involve population genomics (38%) or comparative 226 genomics (27%) (Figure 1D) (data from a questionnaire to species ambassadors, done by 227 ERGA's Data Analysis Committee, DAC, in the framework of Mc Cartney et al. (2023)).

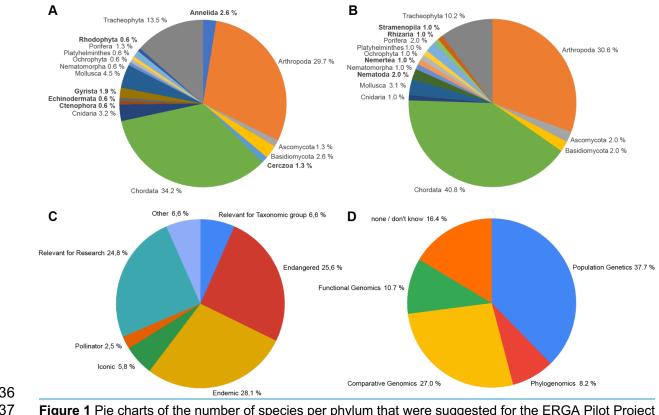
228 Regarding inclusiveness, of the 18 Widening countries represented in the ERGA council 17 had 229 at least one species included in the final list of generated reference genomes. The representation 230 of ITC (Inclusiveness Target Countries) and Widening countries with 44% and 50 % of the 34 231 countries suggesting species is good overall good. However, only 36% or 42% of

- 232 the final species came from ITC or Widening countries, respectively.
- 233

234 Table 2 Feasibility criteria scoring for species suggested as sequencing targets of the ERGA Pilot Project

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Category	1	2	3		
Genome size	<1Gb	1-3Gb	>3Gb		
Sample Availability	Until end April 2020	May-June 2020	July 2020 or after		
Sample Preservation	Freshly collected, flash frozen, -80°C, no preservative, never thawed	in-between 1 and 3 (to be evaluated by sequencing centre)	Not freshly collected and/or thawed several times, and/or not kept in -80°C		
Sample Size	>1g	100mg-1g	<100mg		
Already extracted or taxon known to work well (e.g., vertebrates)		Not tested and not known for the taxon (can be checked with sequencing centres)	Inhibitors known to make DNA extraction and/or sequencing very challenging		
Voucher & SpeciesID	Voucher kept in collection and no ambiguity in species identification		No voucher and/or ambiguous species identification		
Sampling Permits Yes or Not needed (documentation required either way)		Pending	No when needed or No documentation		
Export Regulations	No restrictions between countries where sample will be handled or entire sequencing performed within country	Indexed to conservation status or Nagoya regulations to be clarified	No possibility for obtaining needed permits		



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237 Figure 1 Pie charts of the number of species per phylum that were suggested for the ERGA Pilot Project 238 at the beginning (A) and that are on the list of genomes realised or in production as of April 25th 2023 239 (B). The phyla are indicated together with the percentage of species per phylum. Phyla, which are 240 different between A and B, are highlighted in bold. Additionally, the criterion for choosing the species (C) 241 and the planned downstream analyses (D) are provided in percentages. 242

Ш. FAIR and CARE principles, Metadata Collection and 243 Brokering 244

FAIR and CARE principles 245

246 As the number of initiatives working towards complete reference genomes for all of eukaryotic 247 life are increasing, so too is the demand for freshly collected, wild specimens. This provides an 248 opportune and pertinent moment to revisit biodiversity genomic metadata standards to ensure 249 they are both scientifically comprehensive and also align with current ethical, legal and social 250 standards for data governance. Ensuring that data are findable, accessible, interoperable and 251 reusable (FAIR) is fast becoming a central dogma of the biodiversity genomics community 252 (Wilkinson et al. 2016)¹. Throughout the metadata standard development process (see next 253 section), SSP intentionally and carefully aligned all ontologies to the FAIR principles to safeguard

¹FAIR was introduced by Wilkinson et al. (2016), which has since been accessed 580,000 times and cited 5,636 times

that all ERGA data would have a maximised scientific potential, increased re-usability, and greater longevity.

256 Indigenous Peoples and Indigenous knowledge systems have, and continue to be, treated as 257 subordinate and outside of western science, specifically when considering contextual metadata 258 (Turner 2022). This has had the systematic consequence of severing the connection between 259 Indigenous Peoples and Local Communities with their samples and data. To mitigate the 260 manifestation of this exclusion within ERGA, SSP developed new metadata ontologies to 261 support the disclosure of Indigenous rights and interests by Indigenous Peoples by sample 262 providers. This purposeful inclusion and recognition of Indigenous Peoples and their rights 263 actualises the CARE principles of Indigenous data governance (Carroll et al. 2021) whilst simultaneously working in complementary fashion to the FAIR principles. By creating this space 264 265 at the entry point into ERGA processes, i.e., sample provisioning, SSP provided an opportunity 266 for Indigenous Peoples and knowledge systems to permeate throughout the process of 267 reference genome production and beyond (Figure 2). By operationalizing the FAIR and CARE 268 principles across the metadata ontologies developed, ERGA members are supported to 269 responsibly and openly share data.

270 ERGA Manifest for Metadata Collection and Brokering

271 Developing consortium-wide procedures for metadata collection is an opportunity to set a 272 minimum standard of excellence, and ensures consistency across datasets. This approach is 273 also a challenge since an unintentional exclusion of an important metric will lead to its systematic 274 erasure from all data produced by the consortium. To support ERGA's sampling process, SSP 275 implemented the consortium's first metadata standard, the ERGA manifest, and its supporting 276 documentation (standard operating procedure (SOP)). This SOP and manifest were built on pre-277 existing standards that were developed for an established reference genome production 278 initiative, Darwin Tree of Life (Lawniczak et al., 2022; Shaw et al., 2022), which followed the 279 Darwin Core standard. The manifest supports ERGA's goal to collect standardised, high-guality 280 metadata that remains linked to the genome across the relevant repositories. The highly detailed 281 SOP facilitates completing the ERGA manifest by the Genome Team lead who is responsible to 282 provide information on: 1) sample identifiers (e.g., field and tube numbers, Genome Team lead), 283 2) taxonomic details, 3) sample type (e.g., life stage, organism part), 4) the sequencing partner, 284 5) sample collection event, 6) taxonomic identification and uncertainty, 7) sample preservation, 285 8) DNA barcoding, 9) biobanking and vouchering, 10) regulatory compliances including 286 Indigenous rights and traditional knowledge, and 11) other relevant comments from the Genome 287 Team representative.

- The SOP explains every data point asked for, links to explanatory resources such as tutorial
 videos, and help contacts.
- 290 Expert members of SSP, i.e., sample managers, help genome teams upon request with filling in
- 291 metadata fields and choosing appropriate terms in case of doubt. Sample managers can also
- 292 <u>check investigate manifests prior to submission to avoid frustrating periods of trial and error for</u>

293 <u>sample providers. Based on continuous user feedback, the SOP is updated twice a year under</u>
 294 <u>constant revision to facilitate metadata collection for genome teams.</u>

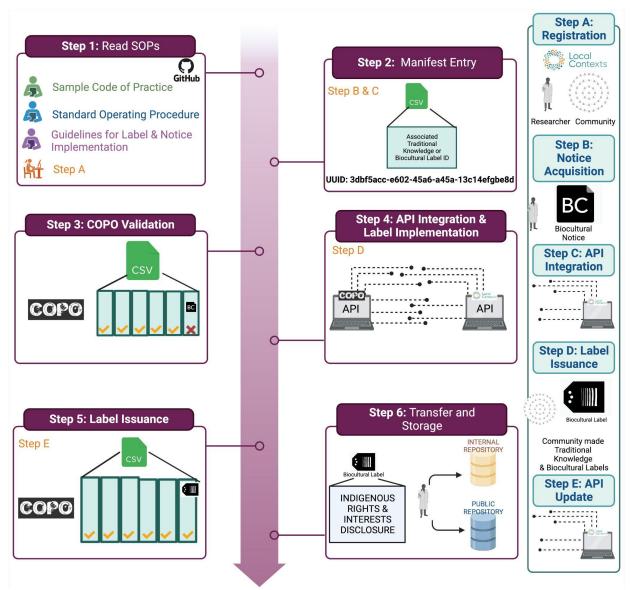
Upon upload of the manifest through the metadata brokering platform <u>COPO</u> (Shaw et al., 2020).
 metadata fields are validated against predefined standards and checklists to ensure terms and
 formats meet both ERGA and data repository expectations. <u>Guidance to this process is provided</u>
 through a visual guide on the COPO help webpage.

Upon manifest validation by the sample managers <u>(part of SSP)</u>, an indicated set of mandatory
 metadata fields are brokered to the <u>European Nucleotide Archive</u> (ENA) under a dedicated
 <u>BioSample</u> entry ultimately connecting the digital sequence data to standardised sample
 metadata.

303 To mitigate the risk of missing information important to specific taxonomic groups or habitats due 304 to own bias (see below), SSP included diverse team members when developing the manifest 305 and planned for bi-annual updates of the metadata protocol so that accidental exclusions could 306 be fixed in a timely manner and allow for sufficient implementation and testing time for front- and 307 backend development. AnySuch and other issues with the manifest encountered by the 308 community can be raised in the ERGA manifest GitHub or by contacting the SSP directly. The 309 ERGA Pilot allowed the SSP committee to test the ERGA manifest on a broad variety of 310 organisms by a pan-European network of researchers. Guidance for understanding and 311 implementing the collection of metadata and vouchers was extensively requested and provided 312 by SSP members. Finalisation of the ERGA manifest and its SOP was achieved through 313 discussions with other ERGA committees, especially ELSI, and the ERGA coordination. The 314 ERGA metadata collection is a semi-automated process that is highly scalable, preparing ERGA 315 for an anticipated increased sample workflow. Validation of the sample manifest is the 316 checkpoint of transitioning to the sequencing workflow. In addition, the SOP and manifest are 317 under constant critical revision based on user feedback aiming to simplify the collection process. 318 The SSP data collection process links biological material, metadata, and sequence information

in a maximally automatised fashion over open access databases and throughout the genome workflow from collection through nucleic acid extraction, sequencing, assembly and annotation steps. While open access genomic information is already a highly appreciated resource, comprehensive metadata enhances its value by making it more reusable. It is crucial that the

323 metadata, sample(s), and derived sequence data are linked from the outset, because the 324 opportunity to link them declines substantially with time (Crandall et al. 2022).



325

326 **Figure 2** ERGA's Biocultural and Traditional Knowledge Labels and Notices implementation protocol.

327 Status Quo of metadata collection amongst biodiversity initiatives

To gain an understanding of the diversity and interoperability between the various metadata collection procedures being implemented within the community, SSP conducted a survey across global biodiversity genomics projects (Figure 3). A total of 24 initiatives that are actively generating high-quality reference genomes for non-human species responded. spanning Africa, North America, Oceania, Europe and Asia^{2*}.

² Notably, the lowest amounts of survey responses were obtained from Asia (the authors note that this is certainly due to our inability to identify appropriate contact points and does not reflect a lower number of biodiversity projects in this continent)

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California Conservation Genomics	0	0	0	0	0	ō	ō	0	0		0				
Vertebrate Genomes Project	0	0	0	0 0 0	0	0	0	0			0		0		
Genomic Observatories Metadatabase	0	0	D	0		0	0	0	0		1				North America
Global Genome Initiative	0	0	D	0	0		0	O		0	0	0	D	0	
Yoder Lab	0	0	O	000	0 0 0		O	0	0	0		1. 15	0		
Beenome100 Project	0	0	0	0	0	0	0	0	0		0	0	P	0	
Natural History Museum of LA County	10122	0	0	0	0	0	0	0	0	0	0	0	0	0	
The Ira Moana Project	0	0	O				0	0					O		
Aotearoa Genomic Data Repository	0	0	O				O	0					D		
Diversity of the Indo-Pacific Network	0	0	O				0	O					D		Oceania
Threatened Species Initiative	0		O	0	0	0	o	0	0	12	E				10101101000
Bioplatforms Australia	0	0	O	0	0	0	0	0	0	2	D	0	0		16 - C
Earlham Institute	0		D					0			0				
Darwin Tree of Life	0	0	0	0	0	0	a	0	0	0	0		D		10000
Aquatic Symbiosis Genomics	0	0	O	0	0	0	O		0	D	0		D		Europe
InvertOmics	0		0				0	0					O		11222202
Catalan BioGenome Project			0	0	0	0	0	0	0	0	O	0	Sec. Co.		N.
Information Category	Proce	dure		Speci	es Informati	lon		Sampl	e Handling	Link to o	ther Geneti	c Resources	Reg	ulatory	

Figure 3 Results summary from the metadata survey conducted across 24 biodiversity initiatives
 worldwide. Red circles within a cell indicate presence, and empty cells indicate absence.

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349 The results indicate that overall, 83% of responding initiatives have a standardised metadata 350 collection procedure in place and 67% have an associated SOP to support and guide 351 researchers in the metadata submission process. In terms of species-specific metadata 352 collection, initiatives prioritise the collection of taxonomic (100%), collection information (96%), 353 biological information (75%) and tissue preservation (75%) over providing more fine-grained 354 information on the taxonomic uncertainty or risks associated with the species being sampled 355 (59%). Almost all initiatives (96%) collected unique specimen and tube/well identifiers as well as 356 the associated principal investigators whereas just 67% required information about the 357 sequencing facility.

The amount of metadata collected about other associated genetic resources from the species sample was relatively low. For instance, only 55% of the 20 projects collect DNA barcoding information within their metadata. Further, just 65% of initiatives collect vouchers and 33% collect cryopreserved samples and require this information as part of their standard metadata collection processes. Finally, 42% of initiatives required some kind of disclosure of regulatory compliance and just 33% of projects required metadata concerning associated Indigenous rights and interest.

365 Scaling Legal Compliance

SSP also focussed on creating an infrastructure that supports and promotes legal as well as ethical and scientifically sound sample collection. As an initial safeguard, SSP supported ERGA to develop a document of best practices for ethical and legal sample collection (ERGA Code of <u>Conduct</u>). All researchers participating in the Pilot were required to agree to these practices in advance of making their metadata manifest submission. These practices detailed expectations surrounding local, regional, national, and international permitting in addition to how to ethically collect samples to minimise harm.

Further, the ERGA manifest contained seven metadata fields regarding the regulation and permit requirements for each sample. <u>These questions comprise comprehensively all permit forms that</u>

375 could be required to obtain a sample for genome sequencing: i) initial question if regulatory 376 compliance is required and adhered to, ii) Applicability of traditional knowledge or biocultural 377 rights with subsequent collection of rights definition, project ID provided by the Local Context 878 Hub and contact information iii) Request for ethics permit applicability, definition and permit iv) 379 Request for sampling permit applicability, definition and permit and v) Request for Nagoya 380 Protocol permit applicability, definition and permit. This comprehensive request for applicability 381 and documentation of compliance raises awareness also for sample providers to respect all 382 regulations.

In partnership with COPO, ERGA required the mandatory upload of permits during the manifest submission process. Expert personnel within ERGA were alerted when a permit had been uploaded into the directory and, where possible, confirmed the appropriate permits had been obtained.

387 The importance of vouchers for biodiversity genomics

388 Voucher specimens in natural history collections are benchmarks against which we compare the 389 world around us. They illuminate how the world has been changing, and especially how we have 390 been changing the world. Reference genomes are a new benchmark. Vouchering is critical to 391 genomics because it provides a permanent, verifiable, and accessioned record of the identity of 392 the organism being sequenced and, in some cases, a sample of its genetic material (biobanking). 393 When determining which of the many available vouchering methods is most appropriate, 394 consideration should be given to e.g., the taxon, its size, its conservation status (Table 3). The 395 SSP determined that a sample voucher helps contextualise the biology of the organism and thus 396 increases the probability that the sequencing data generated will be aligned with FAIR principles 397 aligned and useful into perpetuity.

A driving rationale for vouchering is the fluid nature of taxonomy, as new scientific insights lead to changes in the classification of species. As this happens, the prescribed identity assigned to a sequenced individual could be questioned. In such cases, the presence of a voucher can be used to re-examine the species to confirm, or alternatively revise and update, its identity. Furthermore, vouchers can improve data quality assurance, reduce the risk of data corruption, and eliminate the propagation of confusion when a taxonomic revision has taken place.

Even for taxonomically stable groups, a voucher specimen provides the possibility to join morphological and genome sequence information and verifies the specimen/ species from which the genome was produced. A physical voucher can also be used for other analyses, including photographic, x-ray, CT imaging, and/or chemical analyses such as stable isotopes. A biobanked sample could unlock opportunities for future exploration (e.g., RNA, secondary genetic marker analyses such as methylation). **Table 3** Vouchering methods available to specimens destined for genome sequencing. Note that

411	multiple voucher types may be made for a single genome.
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Desirability Voucher type		Description Suitable for		Potential Issues		
High	Primary voucher	Whole organism is preserved and deposited in a permanent collection. Vouchers can be dried, in a preservation liquid (ethanol), or frozen (e.g., biobanked tissue or cell culture vouchers).	Species that are of a suitable size for a permanent collection (taxon-specific), and can be legally and ethically collected	 Not possible for very large/small species. Species might be too rare to sacrifice for a voucher. Preservation method determines possible additional future uses. 		
	Secondary voucher: to complement - not replace- whole organism vouchering	E-voucher: digital image taken of whole organism and of diagnostic characteristics	Small species requiring destructive sampling to obtain sufficient genetic material for a high- quality genome assembly (e.g., single-cell protist)	 Can require specialist equipment and expertise (e.g., microscope imaging of insect genitalia). May have limited use in taxonomic identification. Diagnostic characteristics may not be known. 		
		Partial Voucher: tissue samples are taken, preserved, curated and stored in permanent collections.	For very large organisms (e.g., a whale), or very small (e.g., small insects), where preservation of the whole organism is not feasible.	 Body part/tissue taker may not represent diagnostic taxonomic characteristics 		
Low		Proxy voucher: a sample that identified as the same species to be sequenced, and was collected from the same time and location	Species that are too small for direct or partial vouchering (e.g., bryophyte)	 May not be the same as the sequenced species 		

IV. Sample provision: connecting genome teams with sequencing centres

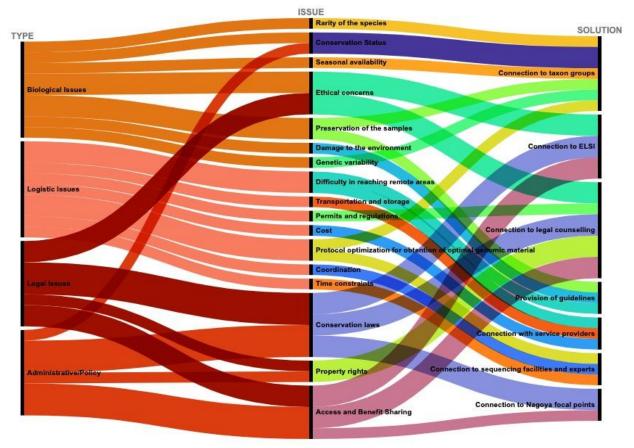
414 Sampling and sample transfer can be a complicated endeavour with its multilayer complexity

arising from three four main categories: biological, logistic, administrative/policy and legal

416 issues. These challenges can strongly influence the outcome of the project and impede the 417 proper transfer of the samples to a sequencing centre (Box 1). The role of SSP is key to

418 overcoming these issues and ensuring the legal, ethical, and timely flow of samples from sample

- 419 collectors to sequencing centres (Figure 4).
- 420



421

- 422 Figure 4 The role of SSP supporting critical issues prior to and after sample collection. Type of issue423 affecting sample provision, description of issues and solutions are indicated.
- 424

425 The distributed genomic infrastructure developed by ERGA promoted and supported the 426 decentralisation of sequencing efforts across Europe. While many sampled species were 427 sequenced within their country of origin, others were shipped to an international sequencing 428 centre. Regardless of the length and duration of shipment involved, ERGA recommended cold-429 chain shipment, which is necessary to preserve the integrity of nucleic acids. Since this can be 430 a challenge for sample providers, ERGA tried to connect sample providers with sequencing 431 centres that were geographically closein proximity and aided in sample transportation within the 432 ERGA network. Maintaining the integrity of nucleic acids This is a prerequisite to meet the EBP

433 standards of genome assembly utilising the current sequencing technology (Dahn et al. 2022). 434 However, samples are often collected in remote locations, where access to appropriate courier 435 service is financially not feasible or simply not available, a challenge that the ERGA Pilot also 436 faced. Further, there is a series of legal procedures that require consideration to ensure 437 compliance with regulations and safety standards, including, among others, chain of custody 438 forms (to document the movement of the samples from collection to sequencing), material 439 transfer agreements (a legal contract between two parties that governs the physical transfer of 440 the biological samples between them, and which establishes the terms and conditions under 441 which the materials will be transferred), import/ export permits (that may be required depending 442 on the country of origin and destination), health certificates (required by some countries to 443 ensure that the samples do not pose a risk to human or animal health), and/or CITES permits 444 (required if the samples are from a species protected under the Convention on International 445 Trade in Endangered Species of Wild Fauna and Flora), as well as ABS/ Nagoya relevant 446 national implementations, among others. The ERGA Pilot project served as an opportunity to 447 understand the magnitude and complexity of these needs and actions in a collective manner, 448 with everyone implicated learning about pieces of information that could make an impact in the 449 success of the full logistics chain. For instance, we learned that different shipping companies 450 operate better in certain geographical regions, and that sometimes it is important to ask them 451 explicitly to refill the dry ice during the transit. We also collectively learned about the bureaucratic 452 idiosyncrasy of each country with respect to export and import permits and Nagoya protocol, 453 with some countries being more flexible and others being more restrictive. All these pieces of 454 information have been shared with SSP and are being leveraged to develop SOPs to facilitate 455 the transit from species collectors to sequencing centres, and will have a strong impact in the 456 implementation of larger projects such as Biodiversity Genomics Europe (see below).

457 Future taxon-specific best-practice guidelines

The biological diversity being sampled by large genome initiatives like ERGA necessitates the development of targeted best-practice sampling guidelines. <u>The approach of having different</u> sampling procedures for different taxa is very commendable as it <u>would</u> eliminates complications arising from structural and functional variations between the taxa.

Such guidelines are imperative <u>to ensure</u> so that sampling efforts minimise the number of samples taken, maximise the data quality, and increase the scientific utility of the sample. To this end, the SSP will take a taxonomic approach that seeks to balance providing a set of guidelines that are comprehensive, with enough specificity to support fit-for-purpose sampling, while simultaneously not providing too much information and materials that may overwhelm field biologists.

To develop these guidelines, separate working groups have been set up for each of the following broad taxon groups: vascular plants, bryophytes and macroalgae, macroinvertebrates, protists, soft bodied invertebrates, fungi and lichens, chordates, and arthropods. The goal of each group is to create a working protocol for the sampling of specimens within that taxonomic group, and those will follow a set structure to ensure consistency and readability. There is a strong

- foundation for these protocols (e.g. <u>dx.doi.org/10.17504/protocols.io.261gennyog47/v1</u>). ERGA
- 474 has the intention of publishing these guidelines in open access over protocols.io

475 A key challenge in developing these guidelines will be to identify and include experts -taxonomic, 476 field, and wet lab biologists- who are willing to voluntarily contribute their time and knowledge to 477 the wider community. The SSP has reached out to the ERGA repeatedly to gain insight into 478 ERGA members' expertise and connect those to SSP. Based on this effort, SSP establishes 479 communication with sample providers and ERGA member institutions that can provide expertise 480 in e.g. sample handling, storing and species identification. This help is provided over the SSP 481 email contact as well as a dedicated channel in the communication platform keybase 482 (https://keybase.io/team/erga.listserv). Vice versa, a future challenge will be to work towards an adoption of these guidelines by the biodiversity community at large. Integrating, documenting, 483 484 and distributing this knowledge and 'know-how' is fundamental to ERGA and its umbrella 485 organisation, the EBP. Based on experiences in the ERGA pilot, members of the SSP and the 486 ERGA BGE project consult with the EBP samples committee and the EBP executive board in 487 areas where ERGA sees a need for larger adoption of processes and standards.

488

489 V. Critical Bias Assessment

490 The biodiversity genomics community is subject to systematic biases that affect the accuracy 491 and completeness of the produced data, and may limit the meaningfulness of the conclusions 492 obtained. Bias comes in many forms, which have different impacts. The ELSI/ JEDI committee 493 is more focused on the human dimension, and the SSP committee focused on country 494 representation and taxonomic biases described here. ERGA as a consortium of European 495 researchers is at its foundation intentionally geographically biased, while at the same time 496 promoting and extending representation and participation of researchers across Europe. In the 497 Pilot, prioritising this aim over the taxonomic breadth of the generated reference genomes 498 resulted in the manifestation of taxonomic biases (see above).

499 Unbalanced representation of genomes being sequenced across the tree of life is 500 common in biodiversity genomics initiatives, causing over-representation of some taxa with data 501 available in public repositories. Non-model organisms and more "difficult" samples remain under-502 investigated because there are few standardised sampling collection, preservation, HMW-DNA 503 extraction, and library preparation protocols available to manage non-optimal situations (e.g., 504 small size, existence of exoskeleton or spicules, presence of substances that impair adequate 505 DNA extraction or sequencing, etc.). This lack of knowledge on certain taxa reflects the available 506 taxonomic expertise. For example, experts in vertebrates, certain arthropod and plant groups 507 are vastly more abundant than for other large taxonomic groups like mollusks, nematodes or 508 annelids (Capa & Hutchings 2021; Engel et al. 2021), which SSP quickly realised while forming 509 taxon expert groups (see above).

510 Bevond taxonomy, other sources of representation bias exist in reference genome 511 projects. Sample bias can occur when samples do not accurately represent the known or 512 unknown heterogeneity of the taxon being studied. SSP encourages sampling from the type 513 locality. Habitat bias occurs when samples are more often collected in certain types of habitats that are more common or more easily accessible, under-representing knowledge about habitat-514 515 specific species (e.g., caves, deep-sea). ERGA aims to target this bias with calls for funded field 516 expeditions to understudied hotspots of biodiversity in Europe. Historical bias can have strong 517 impacts, as samples collected based on prior knowledge or historical information may not 518 accurately reflect the current state of diversity.

519 A prime goal of SSP is to raise awareness of the importance of taxonomic representation for 520 genomics, and biodiversity research more generally, and the study of research deserving 521 groups, species, populations and habitats. SSP has played a key role in creating a bridge 522 between taxonomy- and taxon-specific experts with sequencing centres, and aims to create the 523 conditions to explore the feasibility of genome sequencing for all eukaryotes. Biodiversity 524 genomics benefits the most when it is inclusive in all aspects. Many hotspots of biodiversity exist 525 in Europe, and many are positioned in nations and regions that are deserving of additional support. By creating a European-wide network, SSP aims to support such regions through 526 527 capacity and capability building for genomics.

528

529 VI. Where do we head?

530 We believe that overall, sequencing and assembling the initial cohort of species that entered into 531 ERGA's process was a success story. To a large extent this is thanks to collaboration and 532 alignment with preexisting, well established biodiversity consortia e.g., DToL. Similarly, we hope 533 that our prioritisation efforts, the ERGA metadata manifest, as well as the stewardship of legal, 534 FAIR and CARE information, can be utilised, improved, or adopted by other biodiversity 535 genomics projects, national or international, irrespective of the project size. An immediate 536 example of this is the EU-funded project BGE - Biodiversity Genomics Europe, for which the 537 ERGA initial phase has set the ground for key procedures of the sampling and sample 538 processing process. The BGE consortium unites ERGA with the DNA barcoding community 539 (BIOSCAN Europe) to promote the use of genomics to study and monitor biodiversity and create 540 tools to tackle its decline. BGE will establish ERGA as the European node of the Earth 541 Biogenome Project and formalise coordinated efforts, infrastructures and workflows to generate 542 reference genomes of European species.

543 Towards a balanced and strategic prioritisation of species

As ERGA moves forward, the biases identified are being reflected upon to iteratively improve sampling and prioritisation. As dedicated projects are established, such as BGE, the selection and prioritisation of species for reference genome generation can better approximate governing principles (see above "Selecting species for biodiversity genomics projects"), and be less dependent on circumstantial feasibility aspects and funding availability for particular taxa. These 549 governing principles can be explicitly and objectively included into the species prioritisation 550 process and with a more prominent role, while feasibility will likely remain an important aspect 551 of species selection. Once priorities are established and weighted, the species selection process 552 can be fully automated. Building on the first experiences of ERGA, such a process is being 553 implemented in BGE. This process, which is developed with the larger ERGA community, gives 554 after a check for technical feasibility more weight to taxonomic diversity, country of sample origin, 555 countries with little representation in ERGA and involvesd researchers usingincluding_JEDI 556 criteria (favouringfavoring novel sample providers, -as well-asnd-underrepresented groups, and 557 involvement of non-scientific communities-countries with little representation in ERGA) and 558 applicability of produced genome resource, followed by a check for technical feasibility. ERGA 559 displaying its target species over the platform Genomes on a Tree 560 (https://goat.genomehubs.org/projects/ERGA), in agreement with other nodes of the EBP. 561 ERGA members as well as SSP sample managers engage with other genome initiatives when overlaps are detected and facilitate collaboration in order to prevent parallel efforts. 562

563 A live and comprehensive sampling metadata manifest

564 The ERGA metadata manifest and its SOP are living documents, which are regularly revised 565 under strict version control (https://github.com/ERGA-consortium/ERGA-sample-manifest). 566 During the Pilot phase, it became clear that the metadata core was not entirely comprehensive. 567 For example, the first version could not capture sampling depth and only allowed inputting a 568 precise location. This information is important in the marine context as it was not possible to 569 correctly represent samples from trawls or transects. Updated releases of the manifest have 570 acknowledged these gaps and now comprise fields for e.g., depth and latitudinal and longitudinal 571 coordinates for two points instead of one for sampling transects, extended vocabulary for 572 sampled tissues, etc. As ERGA progresses, adding more extensions might be necessary during 573 the planned regular updates.

574 The guestion that is often raised in regard to metadata collection is what is the trade-off between 575 comprehensiveness versus feasibility. Sampling for reference genome generation has many 576 logistical steps that are important to document in the metadata record. Such extensive collection 577 of metadata appears doable when the emphasis is on single (or a few) representative samples 578 per species while we acknowledge that feasibility and applicability might be different for e.g., 579 population data or already collected material that cannot be obtained again. Yet, as the field of 580 genomics moves forward and technological advances allow extracting more data at higher 581 guality from material with varying guality samples, extending the high ERGA standards to any 582 sample collected for genetic analyses appears as an appropriate perspective. In this light, the 583 increase in frozen archives that ERGA supports will be a treasure trove for genome initiatives.

584 Streamlining legal compliance procedures

585 Biodiversity knows no boundaries and it is blissfully unaware of its traversal distribution across 586 many national, political, and cultural borders that may have varying legal systems. However, 587 ERGA is obligated to respect these borders and the legal systems within, and so a harmonisation of procedures will be a crucial aspect of building a streamlined European sampling infrastructure for reference genome generation. ERGA's network provides cross-country communication, which should be extended to local authorities, and ensure efficient flow of information about specific legal requirements of sampling. Streamlining the steps required to ensure legal compliance therefore is an important way to increase the efficiency of the reference genome generation pipeline.

594 A continued concerted effort

595 Under the umbrella of the EBP and in the light of the progress that sequencing technology and 596 data processing offer, there is a need to scale up the genome generation process. While ERGA 597 has pioneered the establishment of a collaborative transnational effort for reference genome 598 generation in Europe, other regional initiatives advance and face similar challenges. We here 599 call for the establishment of collaborative concerted efforts among different consortia under the 599 EBP flag, unifying standards across the whole workflow, starting with sampling and sampling 601 processing and ending with making data available via open repositories.

602 Glossary

Acronym	Explanation	Ressource			
ABS	Access and Benefit-Sharing	https://absch.cbd.int/			
BGE	Biodiversity Genomics Europe	https://biodiversitygenomics.eu/			
BIOSCAN EUROPE	part of the International Barcode of Life Consortium (iBOL)	https://www.bioscaneurope.org/			
CARE	Collective benefit, Authority to control, Responsibility and Ethics	https://www.gida-global.org/care			
CITES	Convention on International Trade in Endangered Species of Wild Fauna and Flora	https://cites.org			
СОРО	Collaborative OPen Omics	https://copo-project.org/			
DToL	Darwin Tree of Life	https://www.darwintreeoflife.org/			
EBP	Earth Biogenome Project	https://www.earthbiogenome.org/			
DAC	Data Analysis Committee	https://www.erga- biodiversity.eu/team-1/dacdata- analysis-committee			
ELSI	Ethical, Legal, and Social Issues	https://www.erga- biodiversity.eu/team-1/elsi ethical%2C-legal%2C-and-social- issues			
ENA	European Nucleotide Archive	https://www.ebi.ac.uk/ena/browser/ home			
ERGA	European Reference Genome Atlas	https://www.erga-biodiversity.eu/			
FAIR	Findable, Accessible, Interoperable, and Reusable	https://www.go-fair.org/fair- principles/			
GoaT	Genomes on a Tree	https://goat.genomehubs.org/			
ITC	Inclusiveness Target Countries	-			
JEDI	Justice, Equity, Diversity & Inclusion	https://jedicollaborative.com/			
SOP	Standard Operating Procedure	-			
SSP	Sampling & Sample Processing Committee	https://www.erga- biodiversity.eu/team-1/ssp sampling-%26-sample-processing			

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